

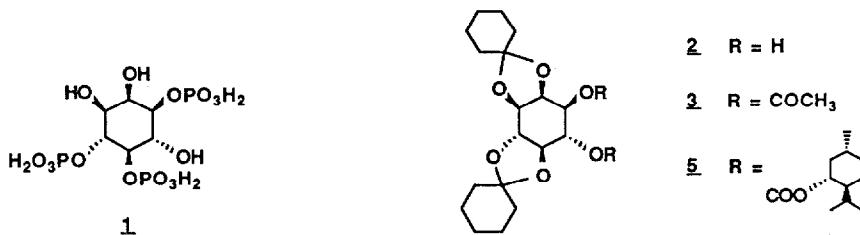
AN EFFICIENT SYNTHESIS OF OPTICALLY ACTIVE D-MYO-INOSITOL  
1,4,5-TRIPHOSPHATE

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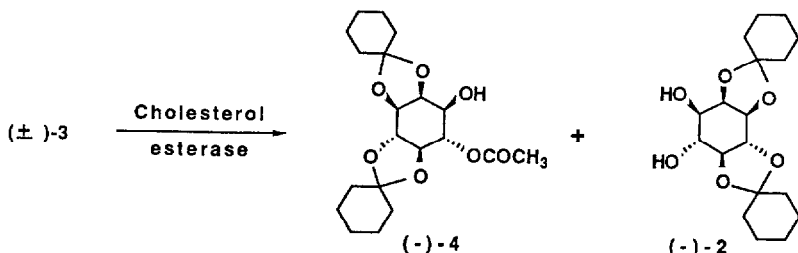
**Summary:** An effective synthesis of D-myo-inositol 1,4,5-triphosphate was developed using a chiral inositide precursor which can be prepared via a facile enzymatic or chemical method.

Recently, much attention has been focused on the metabolic cascade of membrane phosphoinositides, which regulates intracellular  $Ca^{+2}$  mobilization in response to a variety of extracellular signals<sup>1</sup>. This complex information-transducing process is initiated by a receptor-mediated breakdown of phosphatidyl inositol 4,5-bisphosphate with the formation of D-myo-inositol 1,4,5-triphosphate [Ins(1,4,5) $P_3$ ] (1) and 1,2-diacylglycerol, both of which function as second messengers eliciting specific intracellular responses. It has also become clear that Ins(1,4,5) $P_3$  is implicated in the generation of two other biologically active inositol polyphosphates<sup>1</sup>, Ins(1,3,4) $P_3$  and Ins(1,3,4,5) $P_4$ , whose *in vivo* functions still remain unclear. As part of our interest in exploring this intriguing signal-transducing mechanism, we have developed an effective synthetic procedure which allowed the preparation of optically pure 1 in multi-mmol quantities.



In the past<sup>2</sup>, one general problem that has impeded the preparation of enantiomeric 1 in quantities was the optical resolution of inositide intermediates. Cumbersome procedures entailing chiral resolving agents<sup>2</sup> or a chiral HPLC column<sup>3</sup> had to be adopted in the synthesis to separate individual enantiomers in small quantities. Herein, we report a facile synthetic route to optically active 1 using a chiral precursor, (+)-1,2:5,6-di-O-cyclohexylidene-myo-inositol ((+)-2) which can be easily prepared via a facile enzymatic or chemical method.

The diacetate of **(+)-2**<sup>4</sup> (**(+)-3**) was readily digested by a number of hydrolytic enzymes. Of these, cholesterol esterase (Sigma, from bovine pancreas) exhibited high regio- and enantio-preference in removing the acetate functions. In a typical experiment, 1 g of **(+)-3**, dissolved in 2

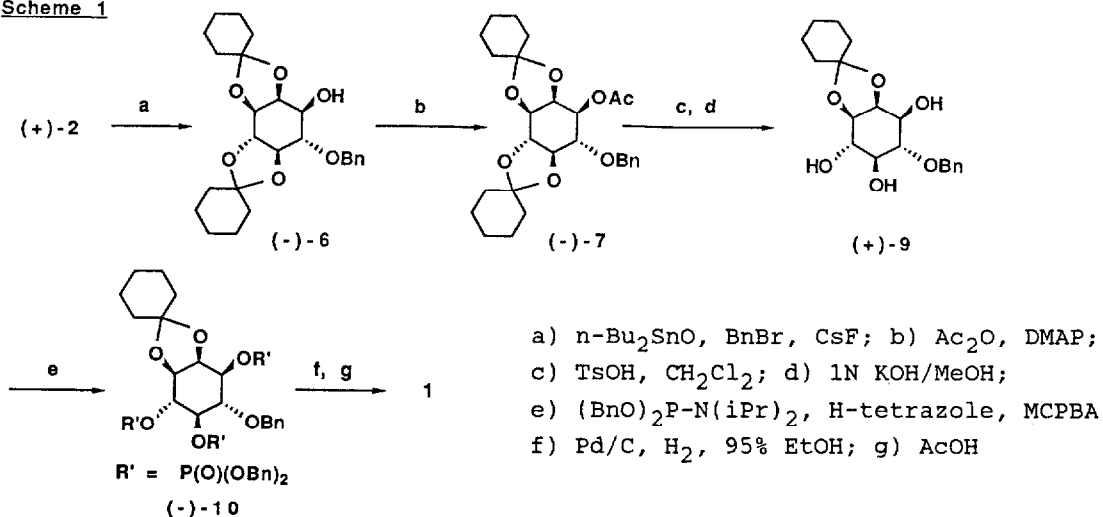


ml of DMF, was finely suspended in 80 ml of phosphate buffer (250 mM, pH 7.5) containing 0.5% Tween 80 and 120 units of cholesterol esterase. The suspension was incubated at 28°C on a rotary shaker (250 rpm) for 168 hr, which yielded a mixture of **(-)-2** (412 mg; 85% *ee*<sup>5</sup>;  $[\alpha]_D^{20} = -15.3$ ,  $c = 1.5$ ,  $\text{CHCl}_3$ ). **(-)-4** (345 mg; 86% *ee*<sup>5</sup>;  $[\alpha]_D^{20} = -8.4$ ,  $c = 1.2$ ,  $\text{CHCl}_3$ ), and a small amount of 3-acetyl-1,2:5,6-di-O-cyclohexylidene *myo*-inositol (30 mg, optical purity not determined). Alkaline hydrolysis of the acetate of **(-)-4** (10 equiv. of 1N KOH/MeOH, 25°C, 1h) afforded **(+)-2** which was subsequently recrystallized from ether-hexane (1:3) to give the diol with optical purity greater than 98% ( $[\alpha]_D^{20} = +18.4$ ,  $c = 1.0$ ,  $\text{CHCl}_3$ ; recovery: 280 mg; combined yield: 70%).

Alternatively, the racemic mixture, **(+)-2**, could also be resolved chemically after being converted to the corresponding di-l-menthyl carbonates<sup>6</sup> (**5a** and **5b**)<sup>7</sup>. The two diastereomers exhibited quite different chromatographic mobility (silica gel TLC (0.25 mm); ether/hexane, 1:3;  $R_f$ : **5a**, 0.87; **5b**, 0.74), and thus were readily separated by  $\text{SiO}_2$  chromatography (ether/hexane, 1:20 to 1:15). Alkaline hydrolysis of the carbonates of **5a** and **5b** gave optically active diols **(-)-2** (>98% *ee*;  $[\alpha]_D^{20} = -18^\circ$ ,  $c = 1.0$ ,  $\text{CHCl}_3$ ), and **(+)-2** (>98% *ee*;  $[\alpha]_D^{20} = +18.4^\circ$ ,  $c = 1.0$ ,  $\text{CHCl}_3$ ), respectively, in nearly quantitative yields. Retrospectively, optically pure **2** may serve as a versatile precursor to various inositol phosphates, which can be illustrated by the efficient synthesis of  $\text{Ins}(1,4,5)\text{P}_3$  (Scheme 1).

**(+)-2** (2.54 mmol) underwent selective partial benzylation by reacting with di-*n*-butyltin oxide (2.8 mmol), followed by benzyl bromide (3.8 mmol) in the presence of cesium fluoride<sup>8</sup> (6.4 mmol) to afford **(-)-6**<sup>9</sup> ( $[\alpha]_D^{20} = -4.2^\circ$ ,  $c = 1.0$ ,  $\text{CHCl}_3$ ) in 91% yield. To prevent the migration of the

Scheme 1



cis-ketal from C-2,3 to C-1,2, which accompanied the direct selective hydrolysis of the trans-cyclohexylidene of (-)-6, the compound was first converted to (-)-7<sup>10</sup> ( $[\alpha]_{\text{D}}^{20} = -14^\circ$ ,  $c = 1$ ,  $\text{CHCl}_3$ ) in quantitative yield. The fully protected inositol (2 mmol) was treated with a catalytic amount of *p*-toluenesulfonic acid (0.2 mmol) in acetone (40 ml) under reflux for 18 min. The resulting compound, without further purification, was subjected to alkaline hydrolysis to give the key intermediate (+)-9<sup>11</sup> ( $[\alpha]_{\text{D}}^{20} = +20.8^\circ$ ,  $c = 1.0$ ,  $\text{CHCl}_3$ ; lit.<sup>2c</sup>  $[\alpha]_{\text{D}}^{20} = +21^\circ$ ,  $c = 0.1$ ,  $\text{CHCl}_3$ ) in 60% total yield (based on 6). Phosphorylation<sup>12</sup> of triol (+)-9 (1.1 mmol) was achieved by the use of *N,N*-diisopropyl dibenzyl phosphoramidite (6.6 mmol), 1-*H*-tetrazole (6.6 mmol) and MCPBA (6.6 mmol) in  $\text{CH}_2\text{Cl}_2$  to afford (-)-10<sup>13</sup> ( $[\alpha]_{\text{D}}^{20} = -4.3^\circ$ ,  $c = 2.42$ ,  $\text{CHCl}_3$ ; lit.<sup>2c</sup>  $[\alpha]_{\text{D}}^{20} = -4.2^\circ$ ,  $c = 0.1$ ,  $\text{CHCl}_3$ ) in 91% yield. Hydrogenolysis of the benzyl groups of (-)-10 (0.9 mmol) ( $\text{H}_2$ , 50 psig, 10%  $\text{Pd/C}$ , 95%  $\text{EtOH}$ , 5 hr) and the subsequent acid hydrolysis of the cyclohexylidene ketal resulted in *D*-myo-inositol 1,4,5-triphosphate 1 (hexasodium salt;  $[\alpha]_{\text{D}}^{20} = -30^\circ$ ,  $c = 0.5$ ,  $\text{H}_2\text{O}$  (pH 9.5); lit.<sup>2c</sup>  $[\alpha]_{\text{D}}^{20} = -30^\circ$ ,  $c = 0.16$ ,  $\text{H}_2\text{O}$  (pH 9.5)) in 90% yield. Thus, enantiomeric 1 was prepared from (+)-2 in 45% overall yield. The  $^1\text{H}$  and  $^{31}\text{P}$  NMR spectra of the synthetic compound 1 were identical to those recently reported<sup>14</sup>.

It should be noted that, in addition to *Ins*(1,4,5) $\text{P}_3$ , enantiomerically pure 2 also provides a facile entry into a variety of optically active phosphoinositides. Currently, syntheses of enantiomeric *Ins*(1,3,4) $\text{P}_3$  and *Ins*(1,3,4,5) $\text{P}_4$  using a similar approach are undergoing in this laboratory.

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#### REFERENCES AND NOTES

1. a) M. J. Berridge, *Ann. Rev. Biochem.*, **56**, 159-193 (1987); b) A. H. Drummond, *Trends Pharmacol. Sci.*, **8**, 129-133 (1987), and references cited therein.
2. Synthesis of optically active **Ins(1,4,5)P<sub>3</sub>**: a) S. Ozaki, Y. Watanabe, T. Ogasawara, K. Kondo, N. Shiotani, H. Nishii, and T. Matsuki, *Tetrahedron Lett.*, **27**, 3157-3160 (1986); b) C. B. Reese, and J. G. Ward, *Tetrahedron Lett.*, **28**, 2309-2312 (1987); c) J. P. Vacca, S. J. deSolms, and J. R. Huff, *J. Am. Chem. Soc.*, **109**, 3478-3479 (1987), and d) C. E. Dreef, R. J. Tuinman, C. J. J. Elie, G. A. van der Marel, and J. H. van Boom, *Recl. Trav. Chim. Pays-Bas*, **107**, 395-397 (1988).
3. S. Ozaki, M. Kohno, H. Nakahira, M. Bunya, and Y. Watanabe, *Chem. Lett.*, 77-80 (1988).
4. Preparation of racemic **2**: P. J. Garegg, T. Iversen, R. Johansson, and B. Lindberg, *Carbohydr. Res.*, **130**, 322-326 (1984).
5. The optical purity of individual inositides was determined by HPLC analysis of the corresponding diastereomeric (+)-MTPA esters using a Whatman Partisil (10 $\mu$ m) column (4.6 mm i.d. x 25 cm). The column was eluted with a solvent mixture of hexane-ether (3:1) at a flow rate of 1 ml/min. Retention times: (-)-**2** di-MTPA ester: 5 min 50 sec; (+)-**2** di-MTPA ester: 7 min 25 sec; (+)-**4** MTPA ester: 8 min 20 sec; (-)-**4** MTPA ester: 8 min 50 sec.
6. The enantiomeric mixture can also be chromatographically separated via the corresponding diastereomeric di-l-menthoxyacetic ester, or the di-S-camphanic ester.
7. Optical rotation: **5a**:  $[\alpha]_D^{20} = -31.8^\circ$ ,  $c = 1.0$ , CHCl<sub>3</sub>; **5b**:  $[\alpha]_D^{20} = -61.1^\circ$ ,  $c = 1.02$ , CHCl<sub>3</sub>.
8. N. Nagashima, and M. Ohno, *Chem. Lett.*, 141-144 (1987).
9. For (-)-**6**,  $\delta_H$  (300 MHz; CDCl<sub>3</sub>) 1.41-1.74 (2H, m), 2.6 (1H, s), 3.48-3.60 (1H, m), 3.90 (1H, t,  $J = 6.6$  Hz), 4.04 (1H, d,  $J = 2.5$  Hz), 4.18 (1H, dd,  $J = 5.4, 7.3$  Hz), 4.35 (1H, t,  $J = 7.3$  Hz), 4.44 (1H, dd,  $J = 3.6$  Hz), 4.73 (2H, ABq,  $J = 11.7, 34.7$  Hz), 7.27-4.41 (5H, m).
10. For (-)-**7**,  $\delta_H$  (300 MHz, CDCl<sub>3</sub>) 1.39-1.65 (2H, m), 2.09 (3H, s), 3.59 (1H, dd,  $J = 6.8, 9.7$  Hz), 3.78 (1H, dd,  $J = 2.0, 5.8$  Hz), 4.01 (1H, dd,  $J = 7.4$  Hz), 4.37 (1H, pseudo t,  $J = 7.4$  Hz), 4.51 (1H, dd,  $J = 3.9, 5.8$  Hz), 4.76 (2H, s), 5.27 (1H, dd,  $J = 2.5, 5.3$  Hz), 7.26-7.36 (5H, m).
11. For (+)-**9**,  $\delta_H$  (300 MHz, DMSO-d<sub>6</sub>) 1.38-1.66 (10H, m), 3.2-3.23 (1H, m), 3.3 (1H, m), 3.46-3.54 (2H, m), 3.75 (1H, m), 3.9 (1H, dd,  $J = 5.7, 7.4$  Hz), 4.25 (1H, dd,  $J = 3.9, 5.3$  Hz), 4.68 (1H, d,  $J = 4.5$  Hz), 4.79 (2H, ABq,  $J = 10.8, 16.8$  Hz), 4.85 (1H, dd,  $J = 4.6, 7.4$  Hz), 7.23 - 7.43 (5H, m).
12. K.-L. Yu, and B. Fraser-Reid, *Tetrahedron Lett.*, **29**, 979-982 (1988).
13. For (-)-**10**,  $\delta_H$  (300 MHz, CDCl<sub>3</sub>) 1.2-1.8 (10H, m), 4.07 (1H, pseudo t,  $J = 8.2$  Hz), 4.2 (1H, pseudo t,  $J = 8.2$  Hz), 4.5-4.60 (3H, m), 4.65 - 5.03 (15H, m), 7.0-7.3 (35H, m).
14. J. C. Lindon, D. J. Baker, R. D. Farrant, and J. M. Williams, *Biochem. J.*, **233**, 257-277 (1986).

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